

factors associated with disease severity could aid in the management and prevention of *S. Aureus* infections.

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## 624. Prevalence of ST171 in *Enterobacter* Isolates from 2001 to 2013 in 15 Hospitals in NYC

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**Background.** ST171 was identified as the most prevalent ST in carbapenem-resistant *Enterobacter* in 26/106 typed isolates from New York City by Gomez-Simmonds et al. There is no study of prevalence of this ST over time. To evaluate a large sample of *Enterobacter*, we designed a PCR assay to identify ST171 isolates rapidly.

**Methods.** Isolates were collected in NYC as part of a cross-sectional Gram-negative antibiotic resistance assessment in the years, 2001, 2003–2004, 2006, 2009, and 2013. Agar dilution MIC were obtained for all isolates as part of these studies. We assayed 284 clinical *Enterobacter* isolates for ST171 using a novel PCR assay, forward primer AGAAGGACGATTGCGCGCGGT and reverse ACTACGGTGGTAAAGAATGATCGCCA. Following amplification, ST171 positive isolates were identified by gel electrophoresis. *Enterobacter* isolates were also assessed for the presence of *bla*<sub>KPC</sub> using a previously described RT-PCR assay.

**Results.** ST171 was identified in 17/284 (6%) *Enterobacter* isolates. This sample collection was heavily antibiotic-resistant with 83/284 *Enterobacter* isolates harboring *bla*<sub>KPC</sub>. Of the 284 isolates, 142 (50%) were resistant to any carbapenem and 113 (40%) were resistant to ceftazidime. Of 17 ST171 positive *Enterobacter* isolates, 14 (82%) were also contained *bla*<sub>KPC</sub>. These isolates were highly resistant, with 10/17 (59%) exhibiting phenotypic resistance to carbapenems and 14/17 (82%) were phenotypically resistant to ceftazidime. Twelve of the 17 isolates occurred in clusters of isolates of the same species occurring in a single hospital at the same time. There were 2 clusters of 3 cases and 3 clusters of 2 patients each. The oldest ST171 strain identified was an *Enterobacter cloacae* isolate collected in 2001. Prevalence of ST171 was calculated for each year of data: 1/40 (2.5%) in 2001, 0/20 in 2003–04, 7/73 (9.6%) in 2006, 6/38 (15.8%) in 2009, and 3/113 (2.7%) in 2013. Each sample collection year included a different group of participating hospitals.

**Conclusion.** Here we evaluate a novel PCR assay for ST171 in *Enterobacter* spp. The oldest ST171 identified from 2001 was carbapenem susceptible suggesting that this strain type later acquired plasmid mediated carbapenem resistance. Prevalence of ST171 tracks with the prevalence of carbapenem resistance with a peak observed in 2009 for both and a decrease in 2013

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## 625. CRISPR-Cas May Prevent Acquisition of Drug Resistance in *Klebsiella pneumoniae*

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**Background.** *Klebsiella pneumoniae* (*Kp*) is a Gram-negative bacterium that causes nosocomial UTIs, pneumonia, and sepsis. Carbapenem-resistant *Kp* (CR-*Kp*) is associated with hospital outbreaks, is difficult to treat, and has high mortality rates prompting study of how resistance is obtained. In U.S. strains, resistance to Carbapenems is primarily conferred by genes *bla*<sub>KPC-2</sub> and *bla*<sub>KPC-3</sub>. Transmission of these genes is via plasmids and to investigate their acquisition, this project analyzed the function of CRISPR-Cas in Carbapenem sensitive *Kp* (CS-*Kp*) hospital strains. The CRISPR-Cas system has been found to suppress homologous gene transfer and prevent integration of new genes by plasmids or bacteriophages. This study's hypothesis is that *Kp* strains that lack CRISPR-Cas can acquire CR plasmids, while those strains that have CRISPR-Cas are protected from gaining these plasmids and can maintain sensitivity to Carbapenems.

**Methods.** *Kp* strains from the urine of patients from Montefiore Medical Center and Stony Brook University Hospital were collected and sensitivity to Carbapenems was determined by chart review. Next, hospital strains were screened for CRISPR-Cas using PCR. 4 CS-*Kp* strains (strains 1 and 2 without CRISPR-Cas, and strains 3 and 20 with CRISPR-Cas) were studied. *bla*<sub>KPC-2</sub>, *bla*<sub>KPC-3</sub>, and control plasmid pPROBEKT-GFP (Km<sup>R</sup>) were then transformed into *Kp* by standard electroporation. Meropenem agar plates for CR-containing plasmids, Kanamycin for control plasmids, and PCR were used to evaluate transformation success.

**Results.** Successful transformation of *bla*<sub>KPC-2</sub>, *bla*<sub>KPC-3</sub>, and the control plasmid was achieved in strains 1 and 2, which lacked CRISPR-Cas. Successful transformation of the control plasmid was achieved in strains 3 and 20. However, neither *bla*<sub>KPC</sub> plasmid could be transformed into strain 3, and while very low success was seen with *bla*<sub>KPC-3</sub> in strain 20, *bla*<sub>KPC-2</sub> could not be transformed into that strain either.

**Conclusion.** This study supports the hypothesis that CRISPR-Cas prevents acquisition of drug resistance plasmids. It is notable that a CS-*Kp* strain with CRISPR-Cas was protected against one KPC gene and not fully against the other. This may indicate a difference in the CRISPR sequences in individual *Klebsiella* strains.

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## 626. Differences in Gene Expression Levels of Methicillin-Resistant *Staphylococcus aureus* Genes Between Persistent and Resolving Bacteremia

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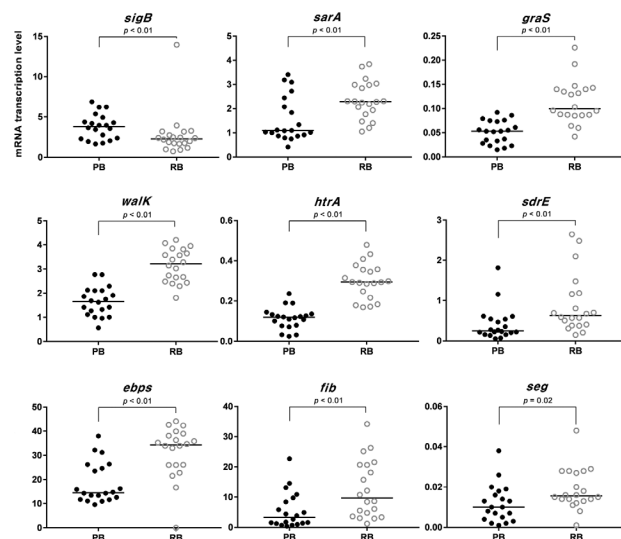
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**Background.** Persistent MRSA bacteremia (PB) is associated with higher mortality than resolving MRSA bacteremia (RB). We previously described that PB and RB isolates had no significant differences in genotypes and microbiologic characteristics. In other small studies, the presence of specific genes or phenotype characteristics was associated with PB, but the results were inconsistent. Aim of this study was to determine whether differences in the expression of major genes contribute to the development of PB.

**Methods.** We analyzed expression levels of major regulatory genes (*agr*, *sarA*, *sigB*, *graRS*, *walKR*, *saeRS*, and *vraRS*) and virulence factors (*htrA*, *prfA*, *murZ*, *spa*, *clfA*, *clfB*, *sdrC*, *sdrD*, *sdrE*, *ebps*, *fib*, *hla*, *psma*, *icaA*, *seg*, and *sen*) in 40 MRSA strains isolated from 20 patients with PB (>7 days) and 20 patients with RB (<3 days) who were matched for clinical and epidemiologic characteristics and bacterial genotypes. Relative gene expression level to *gyrB* was determined using real-time RT-PCR. In the same way, differential expression of the genes between the first and last isolates from 20 patients with PB was analyzed to evaluate changes of gene expression during PB. In addition, RNA-seq was performed on selected MRSA strains to evaluate the overall differential expression of genes.

**Results.** There was no difference in the expression level of *agr* between PB and RB isolates. However, significant differences in gene expression levels between PB and RB isolates were observed in the following genes. Gene with increased expression levels in PB was *sigB*, global regulator. Genes with decreased expression levels in PB included *sarA*, global regulator; *graRS*, *walKR*, and *saeRS*, two-component regulatory systems; *htrA*, *clfA*, *sdrD*, *sdrE*, *ebps*, and *fib*, surface protein genes; *seg* and *sen*, secreted protein genes (Figure 1). A similar trend was found in RNA-seq. There were no significant differences in expression of specific genes between the first and last isolates of PB.

**Conclusion.** Our results suggest that PB may develop due to infection caused by MRSA strain with altered gene expression rather than changes in specific gene expression levels during bacteremia.



**Figure 1. Differences in specific gene expression between PB and RB isolates.** mRNA transcription level is presented as relative expression ratio of target genes to *gyrB*. PB, persistent MRSA bacteremia; RB, resolving MRSA bacteremia.

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## 627. Virulence Factors of Healthcare Associated Infection by Uropathogenic *E. coli* Strains Isolated in Korea

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