



Genetic classification and confirmation of inherited platelet disorders: current status in Korea

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Inherited platelet disorders (IPDs), which manifest as primary hemostasis defects, often underlie abnormal bleeding and a family history of thrombocytopenia, bone marrow failure, hematologic malignancies, undefined mucocutaneous bleeding disorder, or congenital bony defects. Wide heterogeneity in IPD types with regard to the presence or absence of thrombocytopenia, platelet dysfunction, bone marrow failure, and dysmegakaryopoiesis is observed in patients. The individual processes involved in platelet production and hemostasis are genetically controlled; to date, mutations of more than 50 genes involved in various platelet biogenesis steps have been implicated in IPDs. Representative IPDs resulting from defects in specific pathways, such as thrombopoietin/MPL signaling; transcriptional regulation; granule formation, trafficking, and secretion; proplatelet formation; cytoskeleton regulation; and transmembrane glycoprotein signaling are reviewed, and the underlying gene mutations are discussed based on the National Center for Biotechnology Information database and Online Mendelian Inheritance in Man accession number. Further, the status and prevalence of genetically confirmed IPDs in Korea are explored based on searches of the PubMed and KoreaMed databases. IPDs are congenital bleeding disorders that can be dangerous due to unexpected bleeding and require genetic counseling for family members and descendants. Therefore, the pediatrician should be suspicious and aware of IPDs and perform the appropriate tests if the patient has unexpected bleeding. However, all IPDs are extremely rare; thus, the domestic incidences of IPDs are unclear and their diagnosis is difficult. Diagnostic confirmation or differential diagnoses of IPDs are challenging, time-consuming, and expensive, and patients are frequently misdiagnosed. Comprehensive molecular characterization and classification of these disorders should enable accurate and precise diagnosis and facilitate improved patient management.

Key words: Blood platelet disorders, Thrombasthenia, Bernard-Soulier syndrome, Gray platelet syndrome, Platelet storage pool deficiency, MYH9-related disorders

Introduction

There are 2 important components of hemostasis: primary (platelet response to vessel injury and platelet plug formation) and secondary (generation of fibrin deposition by the coagulation cascade).¹⁾ Platelets play a key role in the achievement of adequate primary hemostasis through 4 fundamental mechanisms: adhesion, aggregation, secretion, and procoagulant activity.^{2,3)} Normal platelet adhesion requires the presence of von Willebrand factor (vWF), glycoprotein (GP) receptors on platelets, and the adequate release of platelet granular contents such as adenosine diphosphate (ADP), serotonin, and thromboxane A₂.¹⁾ Thus, familial inherited thrombocytopenia, platelet function disorders, von Willebrand's disease (vWD), or immune thrombocytopenia show similar primary hemostatic problems.⁴⁾

However, the diagnostic confirmation and differential diagnosis of IPDs are complicated, time-consuming, and expensive; thus, many patients with IPD are not appropriately diagnosed.⁴⁻⁶⁾ In most cases, individual megakaryopoiesis, platelet formation, and platelet function are genetically controlled; more than 50 genes associated with IPDs have been identified in recent studies by genome-wide association study and next-generation sequencing.⁷⁾ These genes can be classified according to the specific pathway that is disrupted in platelet formation, e.g., the thrombopoietin (THPO)/THPO receptor (MPL) signaling pathway, transcriptional regulation, granule formation/trafficking/secretion, cytoskeleton regulation, and transmembrane protein signaling pathway (Fig. 1).⁷⁾

Therefore, in this review, representative IPDs based on genetic classification (Table 1) and the current status of these diseases in Korea (Table 2) will be discussed. In particular, this review focuses on IPDs with thrombocytopenia or thrombocytopenia. The diagnostic sequence and flow in IPD is described in Fig. 2.^{4,8)}

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THPO/MPL signaling pathway defects

1. Congenital amegakaryocytic thrombocytopenia

Congenital amegakaryocytic thrombocytopenia (CAMT) (OMIM #604498) is a rare autosomal recessive IPD with severe thrombocytopenia from infancy due to *MPL* gene mutation and

high levels of serum THPO.⁹⁾ The case of a genetically confirmed Korean patient with CAMT harboring a novel mutation in the *MPL* gene was recently reported.¹⁰⁾ CAMT is characterized by megakaryocytopenia in the bone marrow, thrombocytopenia with normal platelet size, and progressive bone marrow failure to aplastic anemia.⁹⁾ Approximately 10%–30% of patients with

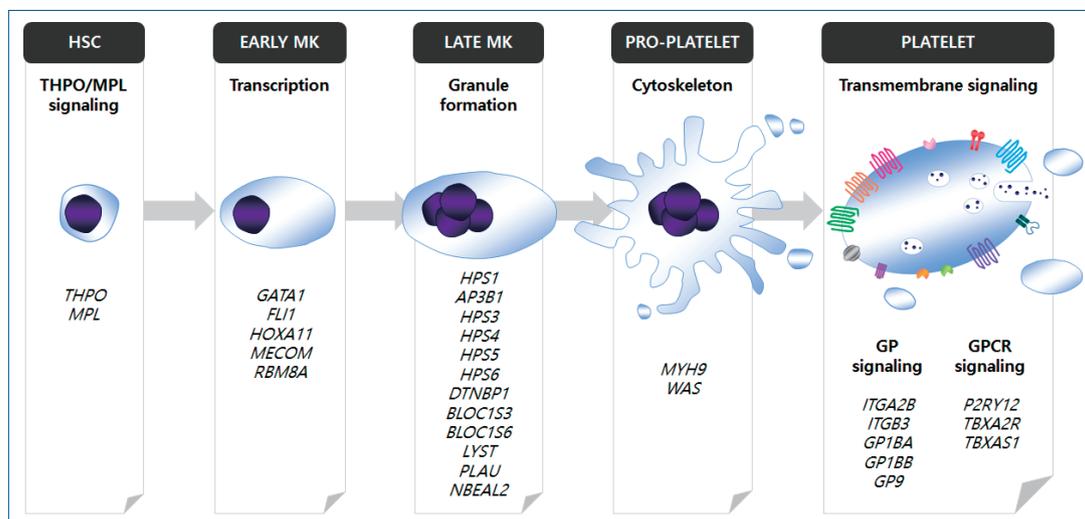


Fig. 1. Identified genes associated with inherited platelet disorders. Each of the genes can be classified according to the specific pathway that is disrupted in megakaryopoiesis or platelet formation. HSC, hematopoietic stem cell; MK, megakaryocyte; GP, glycoprotein; GPCR, G-protein-coupled receptor. Adapted from Lentaigine et al., Blood 2016; 127: 2814–23.⁷⁾

Table 1. Genetic classification of inherited platelet disorders by megakaryopoiesis or platelet formation

Process of platelet formation	Gene	Locus	Protein	Phenotype (OMIM number)	Inheritance	Clinical characteristics
THPO/MPL signaling pathway	<i>THPO</i>	3q27.1	Thrombopoietin	Thrombocytopenia 1 (OMIM #187950) ⁸¹⁾	AD	Familial thrombocytosis Increased megakaryocytes in bone marrow
	<i>MPL</i>	1p34.2	Thrombopoietin receptor	Congenital amegakaryocytic thrombocytopenia (OMIM #604498) ⁹⁾	AR	Absence of megakaryocyte in bone marrow Thrombocytopenia Normal platelet size Elevated serum thrombopoietin Progressive bone marrow failure to aplastic anemia
				Thrombocytopenia 2 (OMIM #601977) ⁸²⁾	AD	Familial thrombocytosis Increased megakaryocytes in bone marrow
Transcriptional regulation	<i>GATA1</i>	Xp11.23	GATA-binding factor 1	X-linked thrombocytopenia with or without dyserythropoietic anemia (OMIM #30036) ^{13–15)}	XR	Thrombocytopenia Large platelet size Variable severity of anemia and hemolysis Dyserythropoietic anemia
	<i>FLI1</i>	11q24.3	Friend leukemia integration 1 transcription factor	Jacobsen syndrome (OMIM #147791) ^{22,23)} Paris-Trousseau type thrombocytopenia (OMIM #188025) ^{17,22)} Chromosome 11q deletion syndrome ¹⁸⁾	AD	Thrombocytopenia Large platelet size Giant α-granules due to fusion of small organelles Dysmegakaryocytopoiesis in bone marrow
	<i>HOXA11</i>	7p15.2	Homeobox protein Hox-A11	Radioulnar synostosis with amegakaryocytic thrombocytopenia 1 (OMIM #605432) ²⁴⁾	AD	Amegakaryocytic thrombocytopenia Radioulnar synostosis
	<i>MECOM</i>	3q26.2	MDS1 and EVI1 complex locus protein	Radioulnar synostosis with amegakaryocytic thrombocytopenia 2 (OMIM #616738) ²⁵⁾	AD	Amegakaryocytic thrombocytopenia Radioulnar synostosis
	<i>RBM8A</i>	1q21.1	RNA binding motif protein 8A	Thrombocytopenia-absent radius syndrome (OMIM #274000) ³¹⁾	AR	Amegakaryocytic thrombocytopenia Absence of radius

Table 1. Genetic classification of inherited platelet disorders by megakaryopoiesis or platelet formation (continued)

Process of platelet formation	Gene	Locus	Protein	Phenotype (OMIM number)	Inheritance	Clinical characteristics
Granule formation, trafficking, or secretion	<i>HPS1</i>	10q24.2		Hermansky-Pudlak syndrome ^{33,34} (OMIM #203300, #608233, #614072, #614073, #614074, #614075, #614076, #614077, #614171)	AR	Normal platelet count Platelet aggregation defect or abnormal response Platelet dense granule defect Oculocutaneous albinism
	<i>AP3B1</i>	5q14.1				Congenital neutropenia
	<i>HPS3</i>	3q24				Pulmonary fibrosis
	<i>HPS4</i>	22q12.1				Granulomatous colitis
	<i>HPS5</i>	11p15.1				
	<i>HPS6</i>	10q24.32				
	<i>DTNBP1</i>	6p22.3				
	<i>BLOC1S3</i>	19q13.32				
	<i>BLOC1S6</i>	15q21.1				
		<i>LYST</i>	1q42.3	Lysosomal trafficking regulator	Chediak-Higashi syndrome (OMIM #214500) ³⁵	AR
	<i>PLAU</i>	10q22.2	Urinary plasminogen activator	Quebec platelet disorder (OMIM #601709) ⁸³	AD	AD Gain-of-function defect in fibrinolysis Thrombocytopenia or normal platelet count Degradation of platelet α -granule contents Platelet aggregation defect or abnormal response
	<i>NBEAL2</i>	3p21.31	Beige and Chediak-Higashi-domain protein	Gray platelet syndrome (OMIM #139090) ⁸⁴	AR	Thrombocytopenia Platelet α -granule defect Large-sized platelets Platelet aggregation defect Gray platelets on light microscopy of Wright-stain Myelofibrosis
Cytoskeleton regulation	<i>MYH9</i>	22q12.3	Nonmuscle myosin heavy chain IIa	MYH9-related thrombocytopenia syndromes (OMIM #155100) ^{37,40-42}	AD	Thrombocytopenia or normal platelet count Large-sized platelets Nephritis Sensorineural hearing loss
	<i>WAS</i>	Xp11.23	WAS protein	Wiskott-Aldrich syndrome (OMIM #301000) ⁵⁰	XR	Thrombocytopenia Small-sized platelets Immunodeficiency Eczema
				Thrombocytopenia 1 (OMIM #313900)= X-linked thrombocytopenia ⁵¹	XR	Thrombocytopenia Small-sized platelets Without profound immunodeficiency Defects of white blood cell cytoskeleton Transient eczema
Transmembrane GP signaling pathway	<i>ITGA2B</i>	17q21.31	GP IIb	Glanzmann thrombasthenia (OMIM #273800) ⁶³	AR	Normal platelet count Platelet aggregation defect
	<i>ITGB3</i>	17q21.32	GP IIIa			
	<i>GP1BA</i>	17p13.2	GP Ib	Bernard-Soulier syndrome (OMIM #231200) ^{68,69}	AR	Thrombocytopenia or normal platelet count Large-sized platelet Platelet adhesion defect
	<i>GP1BB</i>	22q11.2	GP Ib			
	<i>GP9</i>	13q21.3	GP IX			
	<i>GP1BA</i>	17p13.2	GP Ib	Pseudo-von Willebrand disease (OMIM #177820) ^{73,74}	AD	Thrombocytopenia Large-sized platelet Platelet adhesion defect
	<i>GP1BB</i>	22q11.21	GP Ib	22q11 deletion syndrome ⁷⁶⁻⁷⁸ - De George syndrome (OMIM #188400) - Velocardiofacial syndrome (OMIM #192430)	AD	Thrombocytopenia or normal platelet count Large-sized platelet Platelet adhesion defect
GPCR signaling pathway	<i>P2RY12</i>	3q25.1	ADP receptor	Bleeding disorder, platelet-type, 8 (OMIM #609821) ⁸⁵	AR	Platelet aggregation defect
	<i>TBXA2R</i>	19p13.3	Thromboxane A2 receptor	Bleeding disorder, platelet-type, 13 (OMIM #614009) ⁸⁶	AD	Platelet aggregation defect
	<i>TBXAS1</i>	7q34	Thromboxane synthase	Thromboxane synthase deficiency (OMIM #614158) ⁸⁷ Ghosal hematodiaphyseal syndrome (OMIM #231095) ⁸⁸	AD AR	Platelet aggregation defect Platelet aggregation defect
Other	<i>ANO6</i>	12q12	Transmembrane protein 16F	Scott syndrome (OMIM #262890) ⁸⁹	AR	Impaired surface exposure of phosphatidylserine of platelets

AD, autosomal dominant; ADP, adenosine 5'-diphosphate; AR, autosomal recessive; GP, glycoprotein; GPCR, G-protein-coupled receptors; XR, X-linked recessive.

Table 2. Current status of genetic confirmation of inherited platelet disorders in Korea

Phenotype	Genetically confirmed Korean IPD patients	Diagnostic method
Congenital amegakaryocytic thrombocytopenia Jacobsen syndrome	1 Male infant patient reported by Chung et al. ¹⁰⁾	Direct sequencing of <i>MPL</i>
	1 Male premature neonate reported by Noh et al. ¹⁹⁾	Karyotype
	1 Female prenatal case reported by Yoon et al. ²¹⁾	Karyotype through amniocentesis
	1 Female infant reported by Shin et al. ²⁰⁾	Chromosomal microarray and Karyotype
Thrombocytopenia-absent radius syndrome	1 Male infant reported by Kim et al. ²⁹⁾ 1 Prenatal case reported by We et al. ³⁰⁾	Chromosomal microarray Fetal blood analysis
MYH9-related thrombocytopenia syndromes	1 Male adult patient reported by Jang et al. ⁴⁴⁾	Direct sequencing of <i>MYH9</i>
	5 Families (20 patients) reported by Kook et al. ⁴⁵⁾	Direct sequencing of <i>MYH9</i>
	1 Family reported from Lee, et al. ⁴⁶⁾	Direct sequencing of <i>MYH9</i>
	1 Female adult patient reported by Oh et al. ⁴⁷⁾	Direct sequencing of <i>MYH9</i>
	7 Patients (5 male and 2 female) reported by Han et al. ⁴³⁾	Direct sequencing of <i>MYH9</i>
	1 Female pediatric patient reported by Park et al. ⁴⁸⁾ 1 Male patient by K-PHOG study (unpublished data) ^{a)}	Direct sequencing of <i>MYH9</i> Targeted exome sequencing
Wiskott-Aldrich syndrome	1 Male infant reported by Hwang et al. ⁹⁰⁾	PCR-SSCP and direct sequencing of <i>WAS</i>
	1 Male pediatric patient reported by Baek et al. ⁹¹⁾	Direct sequencing of <i>WAS</i>
	2 Male infants reported by Jo et al. ⁵³⁾	Direct sequencing of <i>WAS</i>
	1 Male pediatric patient reported by Kang et al. ⁵⁴⁾	PCR-SSCP of <i>WAS</i>
	1 Male infant reported by Kim et al. ⁵⁵⁾	Direct sequencing of <i>WAS</i>
	2 Families reported by Kim, et al. ⁵⁶⁾	Direct sequencing of <i>WAS</i>
	1 Male infant reported by Lee et al. ⁵⁸⁾	Direct sequencing of <i>WAS</i>
	1 Family reported by Park et al. ⁵⁹⁾	PCR-SSCP and direct sequencing of <i>WAS</i>
	1 Male adolescent patient reported by Yoon et al. ⁶⁰⁾	Direct sequencing of <i>WAS</i>
X-linked thrombocytopenia	1 Male pediatric patient reported by Lee et al. ⁵⁷⁾	Direct sequencing of <i>WAS</i>
	1 Male pediatric patient reported by Yoon et al. ⁶⁰⁾	Direct sequencing of <i>WAS</i>
Glanzmann thrombasthenia	4 Patients reported by Park et al. ⁶⁵⁾	Direct sequencing of <i>ITGB3</i> and <i>ITGA2B</i>
	7 Patients by K-PHOG study (unpublished data) ^{a)}	Targeted exome sequencing

IPD, inherited platelet disorder; K-PHOG, Korean Pediatric Hematology Oncology Group; PCR-SSCP, polymerase chain reaction-single strand conformational polymorphism.

^{a)}These data are in preparation for submission. After targeted exome sequencing, the identified variants were validated by Sanger sequencing.

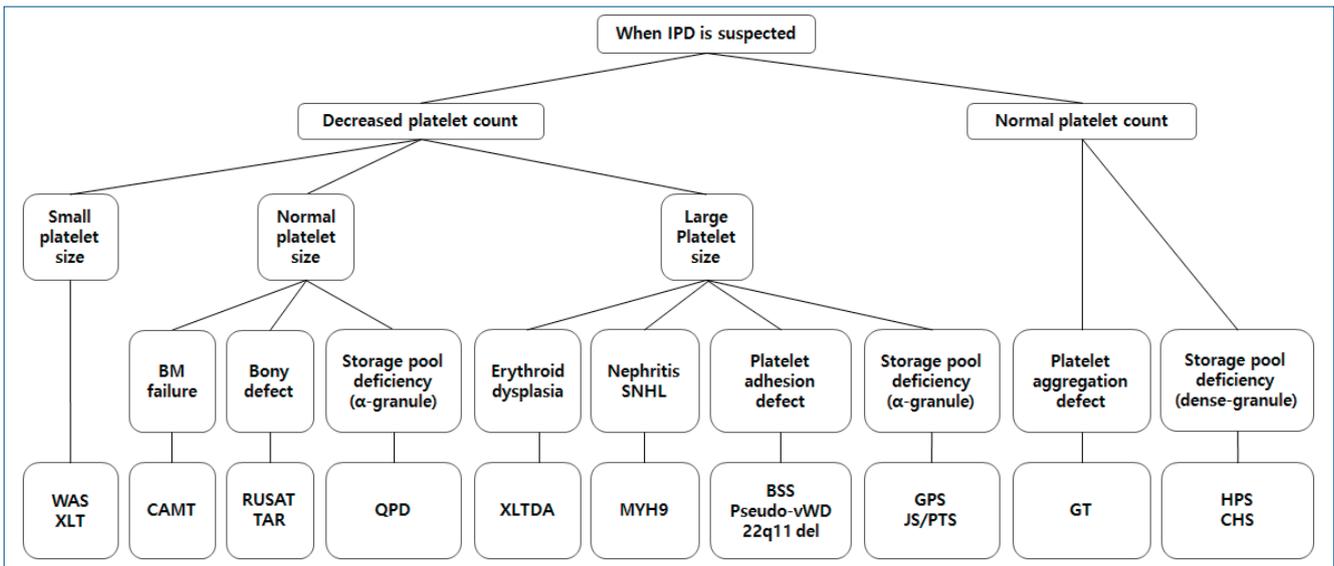


Fig. 2. Diagnostic sequence and flow in inherited platelet disorders (IPDs). BM, bone marrow; WAS, Wiskott-Aldrich syndrome; XLT, X-linked thrombocytopenia; CAMT, congenital amegakaryocytic thrombocytopenia; RUSAT, radioulnar synostosis with amegakaryocytic thrombocytopenia; TAR, thrombocytopenia-absent radius; QPD, Quebec platelet disorder; XLTA, X-linked thrombocytopenia with or without dyserythropoietic anemia; MYH9, MYH9-related disorders; BSS, Bernard-Soulier syndrome; vWD, von Willebrand disease; 22q11 del, 22q11 deletion syndrome; GPS, gray platelet syndrome; JS, Jacobsen syndrome; PTS, Paris-Trousseau syndrome; GT, Glanzmann thrombasthenia; HPS, Hermansky-Pudlak syndrome; CHS, Chediak-Higashi syndrome.

CAMT have orthopedic/neurological abnormalities or intracranial hemorrhage.⁶⁾ The THPO receptor, encoded by the *MPL* gene, promotes megakaryocyte growth and proliferation and may play a role in maintaining hematopoietic stem cells.

^{9,11)} CAMT with bone marrow failure to aplastic anemia is best managed by a hematopoietic stem cell transplantation (HSCT).¹²⁾

Defects in transcriptional regulation

1. X-linked thrombocytopenia with or without dyserythropoietic anemia

X-linked thrombocytopenia with or without dyserythropoietic anemia (XLTA) (OMIM #300367) is a rare X-linked recessive IPD with an unknown prevalence; only a few cases have been reported worldwide.¹³⁻¹⁶ No Korean cases of XLTA associated with *GATA1* have been reported to date. The *GATA1* gene encodes a crucial transcription factor, GATA-binding factor 1, involved in the development of erythrocytes and megakaryocytes in the early stage of hematopoiesis.¹⁴ Depending on the specific *GATA1* mutation, macrothrombocytopenia, variable severity of anemia, hemolysis, and hypercellular marrow with erythroid dysplasia can occur.²

2. Jacobsen syndrome and Paris-Trousseau syndrome

Jacobsen syndrome (OMIM #147791) and Paris-Trousseau syndrome (OMIM #188025) are rare autosomal dominant IPDs associated with the deletion of chromosome 11q.^{17,18} A few Korean cases have been reported.¹⁹⁻²¹ These diseases share several similar phenotypes (dysmorphic features, intellectual disability) and macrothrombocytopenia with giant α -granules due to the fusion of smaller organelles.^{17,22} Chromosome 11q23.3 includes the *FLI1* gene, which encodes an essential transcription factor for megakaryopoiesis.²³ Hemizygous deletion of the *FLI1* gene prevents progenitor cells from undergoing the normal differentiation process, thereby generating a large population of small immature megakaryocytes that undergo lysis.²³

3. Radioulnar synostosis with amegakaryocytic thrombocytopenia

Radioulnar synostosis with amegakaryocytic thrombocytopenia (RUSAT), consisting of RUSAT1 (OMIM #605432) and RUSAT2 (OMIM #616738) types, is a recently identified and introduced autosomal dominant IPD characterized by progressive bone marrow failure and pancytopenia requiring HSCT.^{24,25} Only a few families with these conditions have been reported worldwide,² and no Korean cases have been described in the literature. These syndromes involve amegakaryocytic thrombocytopenia and skeletal defects (limited forearm pronation/supination due to proximal fusion of the radius and ulna).^{24,25} RUSAT1 and RUSAT2 are caused by variants in *HOXA11* and *MECOM* genes, respectively.^{24,25} The protein encoded by the *HOXA11* gene is a DNA-binding transcription factor that regulates gene expression, morphogenesis, and differentiation.²⁴ The oncoprotein EVI1, encoded by the *MECOM* gene, is also a transcriptional regulator involved in hematopoiesis and stem cell self-renewal in humans.²⁶ In mice, *MECOM* protein is expressed at high levels in the embryonic limb bud, suggesting that this gene is also involved in limb development.²⁷

4. Thrombocytopenia-absent radius syndrome

Thrombocytopenia-absent radius (TAR) syndrome (OMIM

#274000), also known as chromosome 1q21.1 deletion syndrome, is a rare autosomal recessive IPD with congenital hypomegakaryocytic thrombocytopenia and bilateral radial aplasia.²⁸ A few Korean cases have been reported to date.^{29,30} A mutation in *RBM8A* gene was recently shown to cause TAR syndrome.³¹ *RBM8A* has several important cellular functions in the production of other proteins by facilitating mRNA transport.³¹ In TAR syndrome, thrombocytopenia becomes less severe over time; the platelet levels reportedly normalized in some cases.^{2,6,28} Some patients also have other congenital anomalies, e.g., lower-limb anomalies, cow's milk intolerance, renal anomalies, cardiac anomalies, intracranial vascular malformation, facial hemangioma, sensorineural hearing loss, and scoliosis.²⁸ A skeletal evaluation and bone marrow study are important in thrombocytopenia such as RUSAT or TAR. Two diseases can emphasize that a thorough orthopedic examination is necessary for patients with thrombocytopenia.

Defects in granule formation, trafficking, or secretion

1. Hermansky-Pudlak syndrome (dense granule disorder)

Hermansky-Pudlak syndrome (HPS) is a rare heterogeneous autosomal recessive IPD group with several genetic defects, including mutations in *HPS1* (OMIM #203300), *AP3B1* (HPS2, OMIM #608233), *HPS3* (OMIM #614072), *HPS4* (OMIM #614073), *HPS5* (OMIM #614074), *HPS6* (OMIM #614075), *DTNBP1* (HPS7; OMIM #614076), *BLOC1S3* (OMIM #614077), and *BLOC1S6* (OMIM #614171).^{7,32} HPS is a disease group of defects in lysosomes, melanosomes, and granule biogenesis.³³ These conditions share features such as oculocutaneous albinism and bleeding symptoms due to platelet dysfunction associated with dense granule defects.^{2,6,32} Owing to the presence of dense granule defects, HPS platelets result in aggregation defects with the absence of second-wave aggregation in response to adrenaline.² Proteins encoded by the *HPS* gene are involved in intracellular vesicle transport, protein sorting, and vesicle docking/fusion.^{2,33} Additional manifestations, including pulmonary fibrosis, granulomatous colitis, neutropenia, or immunodeficiency, can occur according to mutation type.^{2,6,32} No Korean cases of HPS have been reported; however, HPS is a common genetic disorder in Puerto Rico, affecting 1 in 800 individuals, with more than 500 reported cases.³⁴

2. Chediak-Higashi syndrome (dense granule disorder)

Chediak-Higashi syndrome (CHS) (OMIM #214500) is a rare autosomal recessive IPD characterized by ocular cutaneous hypopigmentation, platelet dysfunction, immunodeficiency due to abnormal natural killer cell function, and peripheral neuropathy associated with mutations in the lysosomal trafficking regulator (*LYST*) gene.^{35,36} Thus, CHS disrupts the structure and function of lysosomes and related structures in cells. HPS and CHS share common clinical manifestations (oculocutaneous

albinism and platelet dysfunction due to a dense granule defect) but can be differentially diagnosed by the presence of large peroxidase-positive cytoplasmic granules in neutrophils and more severe symptoms at an earlier age in the case of CHS.³⁵ Few Korean CHS cases have been reported to date.³⁷

Defects in proplatelet formation and cytoskeleton regulation

1. MYH9-related disorders

MYH9-related disorders (OMIM #155100) are autosomal dominant IPDs including May-Hegglin anomaly (MHA), Fechtner syndrome (FTNS), Sebastian syndrome (SBS), and Epstein syndrome (ES).³⁸ All of these syndromes are associated with the *MYH9* gene, which encodes nonmuscle myosin heavy chain IIA (NMMHC-IIA), a component of the contractile cytoskeleton in many tissues, including megakaryocytes, platelets, leukocytes, and the kidneys.^{38,39} The altered distribution of NMMHC-IIA within platelets can be measured by immunohistochemistry or immunofluorescence.³⁹ Thrombocytopenia resulting from defective megakaryocytosis and typical cytoplasmic basophilic inclusion bodies of leukocytes in MYH9-related disorders is caused by the aggregation of abnormal NMMHC-IIA.⁶ According to several studies, MHA, FTNS, SBS, and ES are not separate entities but a single disorder with a wide spectrum of clinical symptoms, e.g., from mild macrothrombocytopenia to the more severe form with sensory neural hearing loss, renal failure, and cataracts.⁴⁰⁻⁴² Platelet counts are typically in the range of $20\text{--}130 \times 10^9/\text{L}$, with elevated mean platelet volume and mild bleeding symptoms.^{6,42} To date, several Korean MYH9-related disorders confirmed by genetic testing have been reported.⁴³⁻⁴⁸

2. Wiskott-Aldrich syndrome and X-linked thrombocytopenia

Wiskott-Aldrich syndrome (WAS) (OMIM #301000) is a rare X-linked recessive IPD, with an incidence of approximately 1–4 per 1,000,000 live male births.^{2,6} Although the prevalence of most IPDs in Korean populations has not yet been determined, the prevalence of Korean WAS is 0.33 per 1,000,000 individuals based on an epidemiological study performed using data of the national registry for primary immunodeficiencies.⁴⁹ The WAS protein (WASP), expressed in hematopoietic cell lineages, has a fundamental role in signal transduction from receptors on the cell surface to the actin cytoskeleton.^{2,6,50,51} Thus, aberrant WASP expression results in defective proplatelet formation and diverse clinical manifestations of WAS, e.g., microthrombocytopenia, eczema, frequent infection due to immunodeficiency, and even malignancy.^{2,6,50,51} Microthrombocytopenia is the characteristics of WAS, and present at birth and varies between 5 and $50 \times 10^9/\text{L}$.² Malignancies occur in 13% of patients with WAS at a mean 9.5 years of age.⁵²

X-linked thrombocytopenia (XLT) (thrombocytopenia 1; OMIM #313900), the milder variant, is characterized by thrombocytopenia with transient eczema and absent or minimal immu-

nodeficiency.⁵¹ WAS gene-associated XLT can be misdiagnosed as chronic ITP because it is milder without frequent infection or eczema.⁶ Several cases of Korean patients with WAS or XLT confirmed by genetic testing have been reported to date.⁵³⁻⁶⁰ The results of hematopoietic stem cell transplantation in Korean patients with WAS have also been reported.^{54,58,61} XLT is difficult to be differentially diagnosed clinically from XLTA (OMIM #300367). In the case of XLT, there may be a family history of WAS, which will help with differentiation between the 2 diseases. In addition, platelet size (large in XLTA, small in XLT) is helpful for differentiating between the 2 diseases.

Transmembrane GP signaling pathway defects

1. Glanzmann thrombasthenia

Glanzmann thrombasthenia (GT) (OMIM #273800) is a rare autosomal recessive IPD with a prevalence of approximately 1 in every 1,000,000 individuals, although a higher prevalence was observed in some populations owing to increased rates of consanguineous marriages.⁶² GT typically develops with loss-of-function variants of the *ITGB2A* or *ITGB3* genes encoding GPIIb and GPIIIa, respectively.⁶³ However, very rare gain-of-function (GOF) variants in the *ITGB3* gene have also been reported, resulting in enhanced fibrinogen binding and severe bleeding.⁶⁴ To date, several Korean cases of GT have been diagnosed clinically or confirmed by flow cytometry; of them, some were further confirmed by genetic analysis.⁶⁵ In patients with GT, there are quantitative and/or qualitative defects in the platelet GPIIb/IIIa complex at binding sites for fibrinogen, vWF, and fibronectin.^{66,67} Thus, patients with GT exhibit normal platelet count/morphology but severely diminished platelet aggregation in response to ADP, epinephrine, serotonin, thrombin, and collagen. Platelet function analyser-100 (PFA-100) measurements are significantly abnormal in cases of GT, with prolonged closure times on ADP/collagen and adrenaline/collagen cartridges.² Impaired platelet agglutination in patients with GT is corrected by ristocetin, which induces the binding of vWF with GPIb/IX.² The application of flow cytometry using antibodies to GPIIb (CD41)/GPIIIa (CD61) is also useful for the diagnosis of GT.²

2. Bernard-Soulier syndrome

Bernard-Soulier syndrome (BSS) (OMIM #231200) is a rare IPD that is typically transmitted in an autosomal recessive manner; its prevalence is estimated to be less than 1 in 1,000,000 individuals.⁶ To date, fewer than 1,000 BSS cases have been reported and studied worldwide.^{2,6,68} No Korean BSS cases have been published to date. BSS is characterized by low platelet counts, typically ranging from less than 30 to $200 \times 10^9/\text{L}$, and the presence of large platelets (macrothrombocytopenia) in a peripheral blood smear.^{6,69} These large platelets are associated with the abnormal development of platelet membranes due to GPIIb abnormalities.⁷⁰ In patients with BSS, platelets have de-

fective surface expression of GPIb-IX-V, which acts as a vWF receptor on platelets, owing to mutations in the *GP1BA*, *GP1BB*, or *GP9* genes.^{2,69,71} As a result, platelets in patients with BSS cannot adhere to the vascular subendothelium and are unable to be agglutinated by ristocetin.⁶⁸ In a study by the International Consortium for BSS, 211 BSS families had mutations in the *GP1BA* (28%), *GP1BB* (28%), or *GP9* (44%) genes.⁶⁸ The closure time in PFA-100 is significantly prolonged in BSS on both cartridges.² The detection of defects in the GPIb-IX-V complex by antibodies to GPIb (CD42b) on flow cytometry is also useful for making the diagnosis of BSS.⁷²

3. Pseudo-vWD

Pseudo-vWD (OMIM #177820), also known as platelet-type vWD or bleeding disorder platelet-type 3, results from GOF variants in the *GP1BA* gene.⁷³ Pseudo-vWD is an autosomal dominant genetic disorder of the platelets, and genetic tests of the *vWF* gene are normal.⁷³ GOF variants in *GP1BA* affect the qualitatively altered GPIb receptor, which then shows increased affinity to vWF.⁷⁴ Accordingly, large platelet aggregates and high-molecular-weight vWF multimers are removed from circulation, resulting in thrombocytopenia and altered vWF multimers. Ristocetin cofactor activity and altered vWF multimers are similar to characteristics of vWD type 2B, a disorder of GOF variants in the *vWF* gene resulting in altered function of the A1 domain of vWF.⁷⁵ In both cases, increased affinity of the GPIb receptor and vWF A1 domain results in thrombocytopenia and abnormal vWF multimers. Thus, DNA analysis is required for differential diagnosis.⁷

4. 22q11 deletion syndromes

The 22q11.21 deletion syndromes, including DiGeorge syndrome (OMIM #188400) and velocardiofacial syndrome (OMIM #192430), may be related to BSS phenotype if the remaining single *GP1BB* gene contains an independently inherited mutation.⁷⁶ These 22q11 deletion syndromes are autosomal dominantly inherited disorders with an estimated incidence of 1 in 4,000 births and are characterized by congenital heart disease, specific facial features, thymic aplasia, frequent infections, cleft palate, hypocalcemia, hypoparathyroidism, learning disabilities, and developmental delay.⁷⁷ If patients must undergo heart surgery because of congenital heart disease associated with 22q11.2 deletion syndrome, physicians should be aware of the serious bleeding risk in these patients.⁷⁸

Conclusions

In the last few decades, researchers have elucidated many of the etiologies of IPDs. The genes involved in a variety of IPDs have been identified, and molecular characterization of these disorders is underway. This information enables a more accurate understanding of IPDs. To enable more accurate and definitive diagnostics, advanced genetic approaches, including next-gene-

ration sequencing, are required for patients with suspected IPD.^{79,80} Although the NGS panel for IPDs has not yet been commercialized and popularized in Korea, interest in IPDs is increasing. As described in Table 2, Korean Pediatric Hematology Oncology Group recently attempted the genetic confirmation of IPDs using targeted exome sequencing as a multicenter study. Although this study has only just started, its findings are expected to be useful for future large-scale studies and the establishment of a Korean IPD registry. It also may be useful for the future development of an NGS panel for IPDs in Korea.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

See the commentary "When to suspect inherited platelet disorders and how to diagnose them" in Volume 63 on page 98.

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