

Histologic Findings and Inflammatory Reactions After Long-term Colonization of *Helicobacter felis* in C57BL/6 Mice

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ARTICLE

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Background: The *Helicobacter felis* (*H. felis*) mouse model has been developed for the research regarding pathogenesis of chronic gastritis and gastric cancer. The aim of this study was to investigate long-term *H. felis* colonization in the stomachs of C57BL/6 mice and subsequent histologic findings and inflammatory reactions including pro-inflammatory cytokines.

Methods: Twenty-three female C57BL/6 mice at 4 weeks of age were gavaged with *H. felis*, and 13 control mice served as vehicle only. The mice were sacrificed at 4, 24, and 52 weeks after inoculation. The infection status and degree of inflammation were determined by culture and histopathology. The level of gastric mucosal myeloperoxidase (MPO), tumor necrosis factor alpha (TNF- α), and interleukin-1beta (IL-1 β) were measured by ELISA.

Results: The overall infection rate was 100%, as determined by the culture and histology. At 4, 24, and 52 weeks, the neutrophil and monocyte scores were significantly higher in infected mice than in control mice. At 24 weeks after inoculation, most of the infected mice showed mucosal atrophy with or without metaplasia, and a few showed focal dysplasia. Adenocarcinoma was observed in one mouse at 52 week post-infection. Gastric mucosal MPO and IL-1 β levels were significantly higher in infected mice than those in control mice at 24 and 52 weeks. However, the expression of gastric mucosal TNF- α was not significantly different between the infected and control mice at any time-point.

Conclusions: Long-term *H. felis*-infection in C57BL/6 mice provoked a severe inflammatory reaction and it progressed into atrophy, metaplasia, dysplasia and cancer. IL-1 β might play an important role in the inflammatory response of mice to *Helicobacter* species. (J Cancer Prev 2014;19:224-230)

Key Words: *Helicobacter felis*, Inflammation, Interleukin-1beta, Tumor necrosis factor-alpha

INTRODUCTION

Helicobacter pylori (*H. pylori*) infection is a major etiologic factor associated with gastric cancer,¹ and the bacterial pathogen was classified as a "definite biological carcinogen" by the World Health Organization in 1994.² Various experimental animal models have been used to elucidate the underlying pathological and biochemical mechanisms of *Helicobacter*-associated gastric cancer. A limited number of *H. pylori* strains have successfully colonized the mouse stomach. The most common organisms that colonize the mouse stomach are *H. pylori* Sydney strain 1 (SS1)

and *Helicobacter felis* (*H. felis*).³ *H. felis*, first isolated from the gastric tissue of the cat, causes natural infections in mice and is analogous to *H. pylori* in humans.⁴ Infection of the C57BL/6 strain of mice with *H. pylori* SS1, a mouse-adapted *H. pylori* strain, results in chronic colonization and hypertrophy, but these mice do not always develop dysplasia and carcinoma.⁵ Chronic *H. felis* infection induces severe inflammation, atrophy, metaplasia, dysplasia, and gastric cancer in C57BL/6 mice.⁶

H. pylori-associated gastric inflammation is mediated by pro-inflammatory cytokines such as the tumor necrosis factor (TNF)- α , interferon- γ , interleukin (IL)-1 β , IL-6, IL-8, and IL-18.^{7,8}

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IL-1 β is an important inflammatory mediator and may be involved in carcinogenesis.^{9,10} TNF- α is also associated with inflammation, immune regulation, and tissue repair and is an important factor in the development of digestive diseases.¹¹

The aim of this study was to investigate chronic *H. felis* colonization in the stomachs of C57BL/6 mice and inflammatory reactions, including the production of pro-inflammatory cytokines. We report the results of these investigations and the subsequent histologic findings.

MATERIALS AND METHODS

1. Animals and *H. felis* infection

Four-week-old female C57BL/6 mice (Orient Co., Ltd., Seoul, Korea) weighing 10-15 g were used for the experiments. All mice were housed in a cage maintained at 23°C with a 12/12-hour light/dark cycle under specific pathogen-free conditions. The mice were administered orogastrically with vehicle only (0.25 mL, for controls) or vehicle containing more than 1×10^7 colony-forming units (CFU)/mL of *H. felis* (ATCC 49179), 5 times every other day. The mice were sacrificed by CO₂ asphyxiation at 4, 24, and 52 weeks after inoculation. All experimental procedures were approved by the Institutional Animal Care and Use Committee (IACUC) of Seoul National University Bundang Hospital (IACUC number: BA1004-059/016-01).

2. Histopathology

At necropsy, stomach tissue was taken from the greater curvature beginning at the squamocolumnar junction and ending at the gastroduodenal junction. Linear gastric strips were fixed in 10% formalin solution, processed by standard methods, embedded in paraffin, sectioned at 5 μ m, and stained with hematoxylin and eosin. The stomach mucosa was histologically examined for inflammatory and epithelial changes and for the presence of *H. pylori*. The degree of neutrophil infiltration, mononuclear cell infiltration, atrophy, and metaplasia was assessed according to the updated Sydney classification as follows: 0, absent; 1, minimal; 2, mild; 3, moderate; 4, marked.

3. Measurement of mucosal myeloperoxidase, tumor necrosis factor alpha, and interleukin-1 beta

Ten milligrams of scraped mucosa were homogenized for 30 seconds with a Polytron homogenizer in 200 μ L of ice-cold lysis buffer (200 mM NaCl, 5 mM EDTA, 10 mM Tris [pH 7.4], 10% glycerin, 1 mM PMSF, 1 μ g/mL leupeptin, and 28 μ g/mL aprotinin). The cell suspensions were centrifuged at 13,000 rpm for 15

minutes, and the resulting supernatant was assayed using a myeloperoxidase (MPO) ELISA kit (HyCult Biotechnology, Uden, The Netherlands). For TNF- α and IL-1 β , the appropriate kits from R&D Systems (Minneapolis, MN, USA) were used following the manufacturer's instructions. Protein concentration was measured using a Bio-Rad Protein Assay Kit (Bio-Rad Laboratories, Hercules, CA, USA). The concentration of each cytokine was measured in picograms per milligram of protein. All assays were performed in triplicate.

4. Statistical analysis

Data are expressed as means \pm SEMs. Comparison of between the 2 groups (experimental and control) was performed using the Mann-Whitney U test or Fisher's exact test. P values less than 0.05 were considered statistically significant. All statistical analyses were performed using SPSS software (version 20.0; SPSS Inc., Chicago, IL, USA).

RESULTS

1. *Helicobacter felis* colonization

The *H. felis* infection status was evaluated by culture or histology. Of the 23 infected mice, 14 (60.8%) showed positive results in culture. However, all infected mice (100%) tested positive for infection by histopathology (Table) and had *H. felis* density scores of 1.5 at week 4, and 1.0 at week 24 and 52 (Fig. 1A). No *H. felis* were isolated in the stomach of the control mice.

2. Histopathology

The neutrophil and monocyte grades of infected mice peaked at week 24 and were significantly higher at all of time points compared with the control mice (Fig. 1B-C). No atrophy occurred up to week 4, however, all infected mice had significantly higher atrophic scores than control mice after 24 weeks. The mean

Table. Histologic findings in mice infected with *H. felis* over 1 year

Histologic findings	Control	Weeks after inoculation		
		4	24	52
<i>H. felis</i> (+)	0/13	4/4	8/8	11/11
Neutrophil	0/13	4/4	8/8	11/11
Monocyte	0/13	3/4	8/8	11/11
Mucosal atrophy	0/13	0/4	8/8	11/11
Intestinal metaplasia	0/13	0/4	5/8	9/11
Lymphoid aggregation	0/13	1/4	7/8	11/11
Hyperplasia	0/13	0/4	5/8	0/11
Dysplasia	0/13	1/4	2/8	3/11
Gastric cancer	0/13	0/4	0/8	1/11

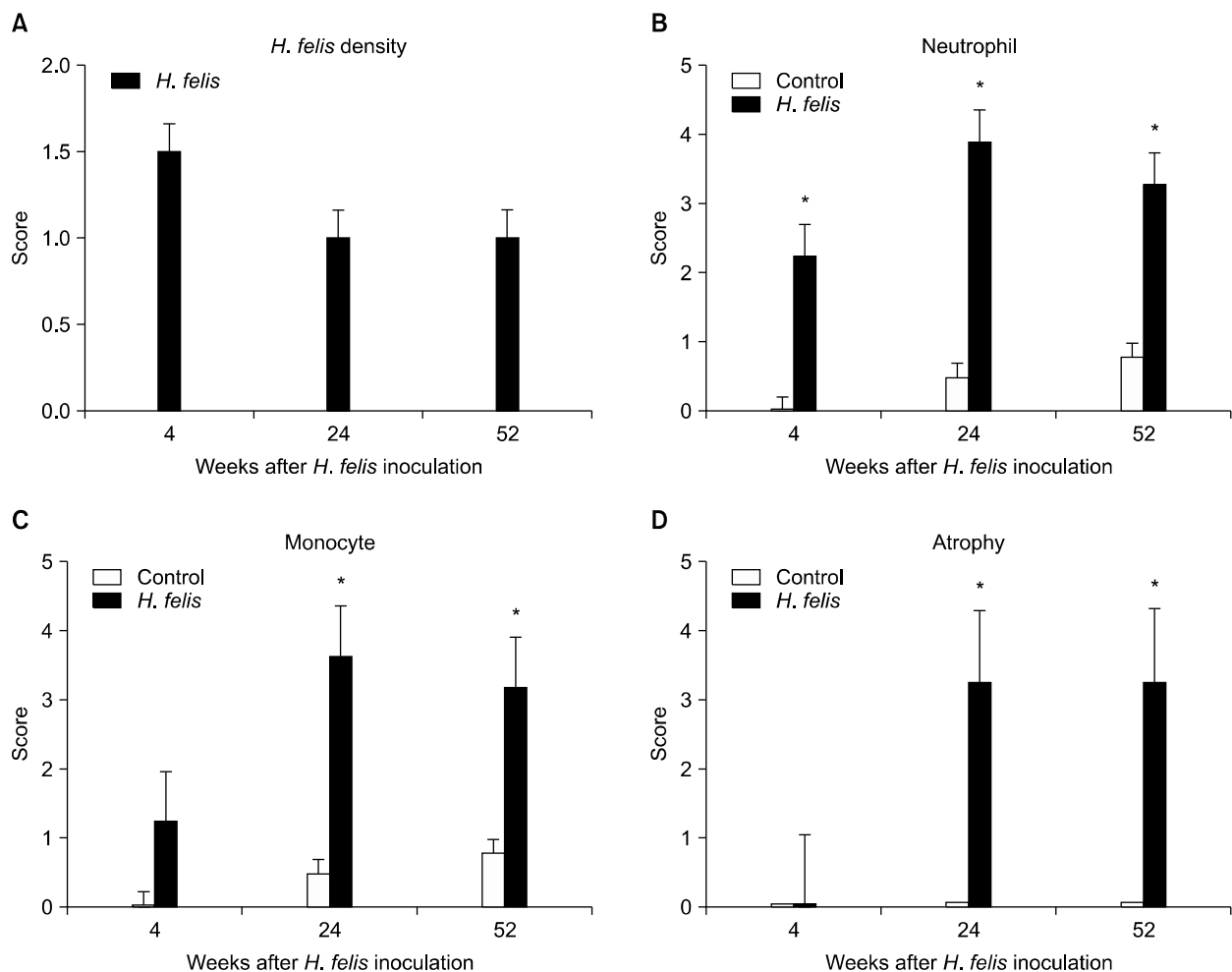


Figure 1. Gastric histopathology scores of *Helicobacter felis* (*H. felis*) density (A), Neutrophil (B), Monocyte (C), Atrophy (D) at 4, 24, and 52 weeks after *H. felis* inoculation. The neutrophil and monocyte grades of infected mice peaked at week 24 and were significantly higher at all of time points compared with the control mice. In case of atrophy significantly higher atrophic scores appeared at 24 weeks and it continued up to 52 weeks. Data are presented as means \pm SEMs. * $P < 0.05$ compared with controls at the same time-point.

atrophy scores were 3.25 in week 24 and 3.27 in week 52 (Fig. 1D).

All control mice demonstrated normal histopathology (Fig. 2A). At 4 weeks after inoculation, neutrophil and monocyte infiltration occurred in all infected mice, suggesting chronic active gastritis (Fig. 2B). In particular, one mouse at week 4 demonstrated lymphoid aggregations, erosion, and high grade dysplasia. At 24 weeks after inoculation, mucosal atrophy and mucous metaplasia appeared in all infected mice. The normal architecture was completely lost and replaced by pseudo-pyloric metaplasia (Fig. 2C) and a hyperplastic change of the mucosa was observed (Fig. 2D). Focal low-grade dysplasia was also noted in some mice. At 52 weeks after inoculation, dysplastic change (Fig. 2E) and adenocarcinoma with focal submucosal invasion (Fig. 2F) was noted in a few mice (Table).

3. Expression of pro-inflammatory cytokines

At 24 and 52 weeks, the gastric mucosal MPO level (ng/mL) in *H. felis*-infected mice was significantly higher (week 4, 11.8 ± 5.2 ; week 24, 36.9 ± 4.4 ; week 52, 35.8 ± 5.1 ; $P < 0.05$) than that in control mice (Fig. 3A). The gastric mucosal IL-1 β expression (pg/mL) in *H. felis*-infected mice peaked at week 24 and decreased trend (week 4, 36.4 ± 12.5 ; week 24, 186.0 ± 15.8 ; week 52, 102.2 ± 8.5). At weeks 24 and 52, the level of IL-1 β expression was significantly higher for infected mice than that for control mice ($P < 0.05$) (Fig. 3B). The gastric mucosal TNF- α expression (pg/mL) in *H. felis*-infected mice (week 4, 11.8 ± 5.2 ; week 24, 36.9 ± 4.4 ; week 52, 35.8 ± 5.1) did not show any significance compared with control mice and there was no correlation between the weeks and gastric mucosal TNF- α level (Fig. 3C).

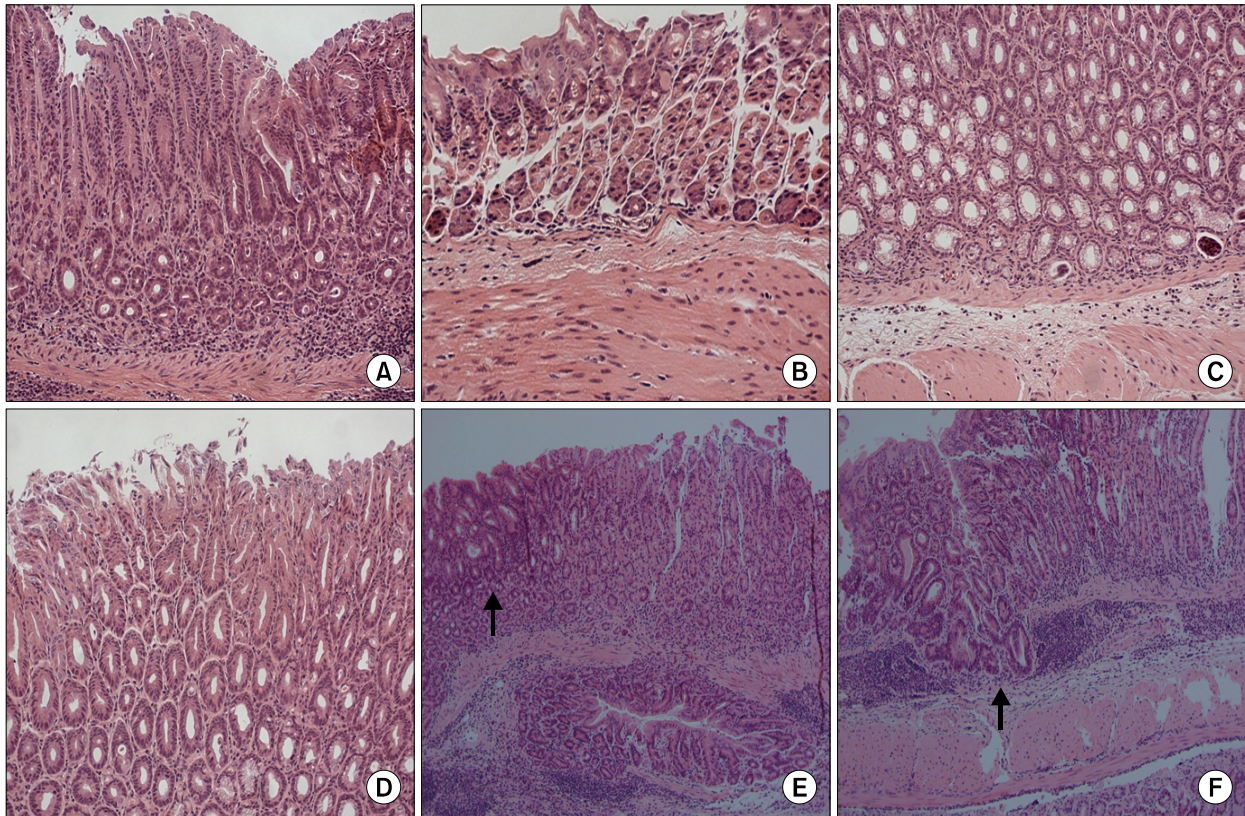


Figure 2. Histopathologic findings of the gastric mucosa at each time-point. Hematoxylin and eosin (H&E) staining; $\times 200$ (A-D), $\times 100$ (E, F). Normal gastric mucosa (A), Marked neutrophil infiltration at 4 weeks after inoculation (B), pseudopyloric metaplasia (C) and hyperplasia (D) at 24 weeks after inoculation, low-grade dysplasia (arrow) (E) and adenocarcinoma with focal submucosal invasion (arrow) (F) at 52 weeks after inoculation.

DISCUSSION

In this study, we observed severe inflammation, glandular atrophy, metaplasia, dysplastic change, and adenocarcinoma in mice stomachs at 4, 24, and 52 weeks after *H. felis* inoculation and successfully established the chronic *H. felis* infection model in C57BL/6 mice. In addition, we analyzed the levels of gastric mucosal pro-inflammatory cytokines such as IL-1 β and TNF- α .

A reliable animal model of *H. pylori* infection is necessary for evaluating vaccine efficacy and for understanding the pathological mechanism of the organism. After discovery of *H. pylori* by Marshall and Warren in 1984,¹² many attempts had been made to establish animal models. In early 1990, *H. felis*, a close relative of *H. pylori* that was isolated from the cat stomach, and it was used because human clinical strains of *H. pylori* were not able to colonize mice stomachs.¹³ *H. felis*-infected C57BL/6 mice showed the classic sequence of histologic changes consistent with human infection, including gastritis, oxyntic gland atrophy, surface epithelial proliferation,

metaplasia, dysplasia and adenocarcinoma.¹⁴ Sakagami et al.¹⁵ reported that C57BL/6 mice infected with *H. felis* showed moderate to severe chronic active gastritis, which increased in severity over time and atrophic changes after 6 months. In other studies of a longer duration, *H. felis*-infected mice showed gastric metaplasia, dysplasia and invasive cancer.^{6,14,16} In 1997, Lee et al.¹⁷ first introduced mouse adapted *H. pylori* strain, "the Sydney strain of *H. pylori* (SS1)". *H. pylori* SS1 shows high levels of colonization in C57BL/6 mice. However, previous studies showed that chronic *H. pylori* SS1 infection in C57BL/6 mice results in chronic colonization and hypertrophy, but these mice do not always develop dysplasia and carcinoma.^{18,19} These differences may be associated with differences in the colonization pattern between *H. pylori* SS1 and *H. felis*. *H. pylori* SS1 tends to localize in the upper foveolae, whereas *H. felis* tends to colonize the deep foveolae and antral glands.^{17,20} In addition, *H. pylori* SS1 adheres to the epithelium at the top of the pits, as observed by electron microscopy. In contrast, *H. felis* has only been observed free in the gastric mucus and does not adhere to the epithelium.^{17,21} We

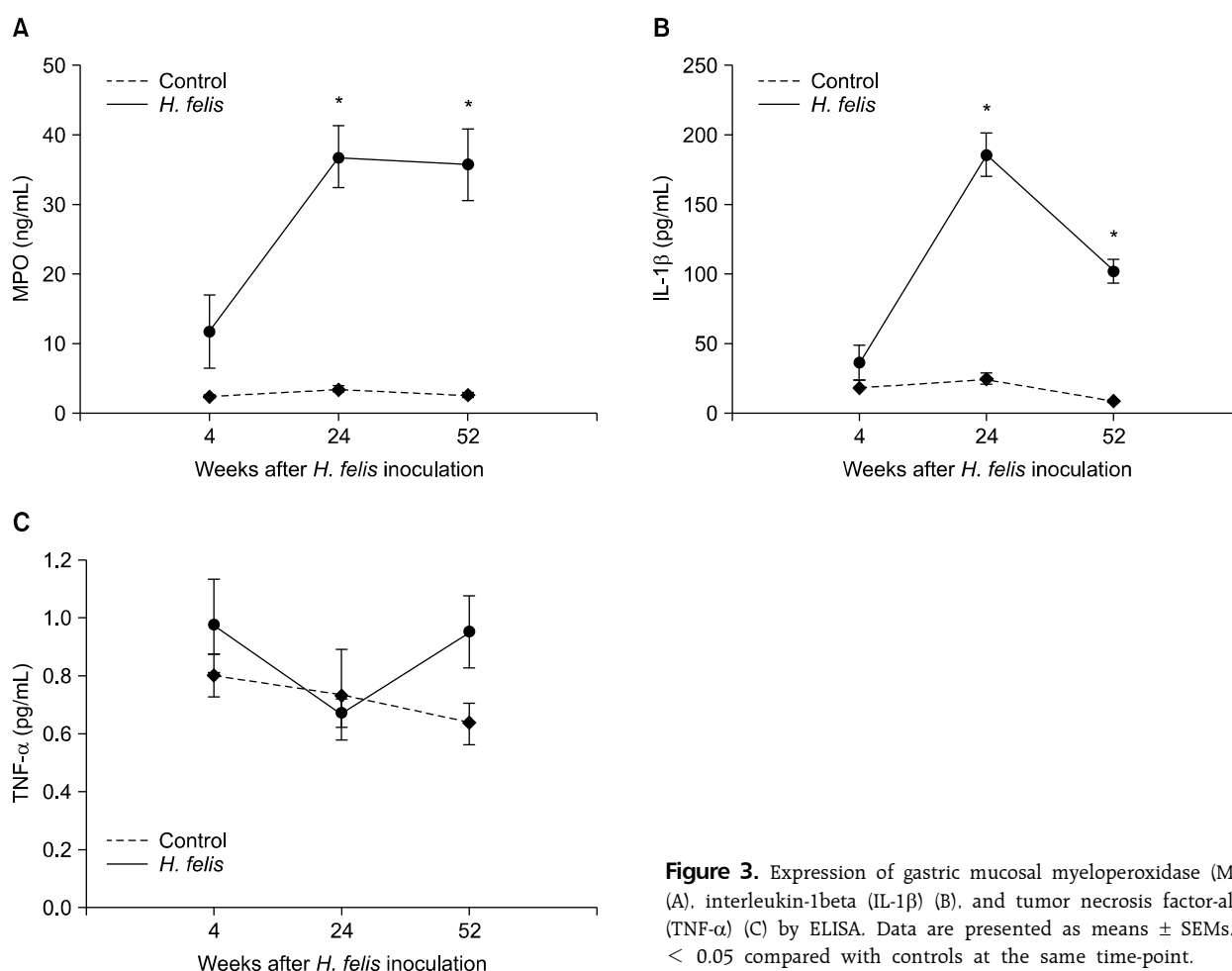


Figure 3. Expression of gastric mucosal myeloperoxidase (MPO) (A), interleukin-1beta (IL-1β) (B), and tumor necrosis factor-alpha (TNF-α) (C) by ELISA. Data are presented as means ± SEMs. * $P < 0.05$ compared with controls at the same time-point.

previously reported about *H. pylori*SS1 infection in C57BL/6 mice with minimal to mild (score 1-2) neutrophil infiltration 4 weeks after inoculation.²² Furthermore, the mean gastric mucosal MPO level was 3.4 ± 0.6 ng/mL at 4 weeks after *H. pylori* infection.²² In the present study, *H. felis* infection resulted in mild to moderate neutrophil infiltration (score 2-3) during the same period, and one infected mice showed high-grade dysplasia 4 weeks after inoculation. The mean gastric mucosal MPO level during same period was 11.8 ± 5.2 ng/mL, and the level then markedly increased more than 30 ng/mL at weeks 24 and 52. Therefore, *H. felis* apparently provoked more severe inflammation than did the *H. pylori*SS1 strain and ultimately led to adenocarcinoma.

Gastric mucosal pro-inflammatory cytokines such as IL-1β, IL-6, IL-8, and TNF-α increase in *H. pylori* infected animal. IL-1β physiologically induced by *H. pylori* infection enhanced gastric carcinogenesis by affecting both inflammatory and epithelial cells.²³ Sun et al.²⁴ reported that there was an increased IL-1β expression at all time-points during long-term *H. pylori* infection.

TNF-α, produced mainly by activated macrophages, is a cytokine with pleiotropic function; editing injured cells, effete immune cells and repairing inflammatory damage.²⁵ The role of TNF-α in experimental mice is not clear; results are inconsistent. Several studies reported no detectable difference in the grade or activity of gastritis in TNF receptor deficient mice compared with wild-type mice after *H. pylori*^{26,27} or *H. felis*²⁸ infection. Our results agreed with those of these studies; the gastric mucosal TNF-α expression in *H. felis*-infected mice was not significantly different from that in control mice. This observation supports the results of our previous *H. pylori* study.²² Therefore, TNF-α does not appear to participate in the development of an inflammatory response caused by *Helicobacter* species. However, in their study on chronic *H. pylori* infection-induced gastritis using Mongolian gerbils, Sun et al.²⁴ reported that the expression of TNF-α increased significantly at 6, 32 and 56 weeks after inoculation. Hasegawa et al.²⁹ reported that the induction of gastritis by *H. felis* infection was clearly depressed in TNF-α knockout mice

compared with C57BL/6 mice and suggested that TNF- α is critical for the induction of gastritis by *H. felis* infection. Further animal experiments are needed to clarify the role of TNF- α in the inflammatory responses of *H. pylori* infection.

In conclusion, *H. felis* well colonized the stomachs of C57BL/6 mice, and provoke severe inflammation, and eventually induced mucosal atrophy, metaplasia, dysplasia and cancer. IL-1 β plays an important role in *H. felis* gastric inflammation; however, TNF- α is not involved in the inflammatory processes. Further trials are needed to clarify these inflammatory mechanisms.

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CONFLICTS OF INTEREST

No potential conflicts of interest were disclosed.

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